

# Lecture B

Exhaustive and non-exhaustive algorithms with  
resampling

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# Overview

B1. Cross validation

B2. [The mplot package](#)

- Artificial example: *Information criteria can be blind*
- Variable inclusion plots
- Model stability plots
- Bootstrapping the lasso

B3. Subtractive stability measures

# B1. Cross-validation for model selection

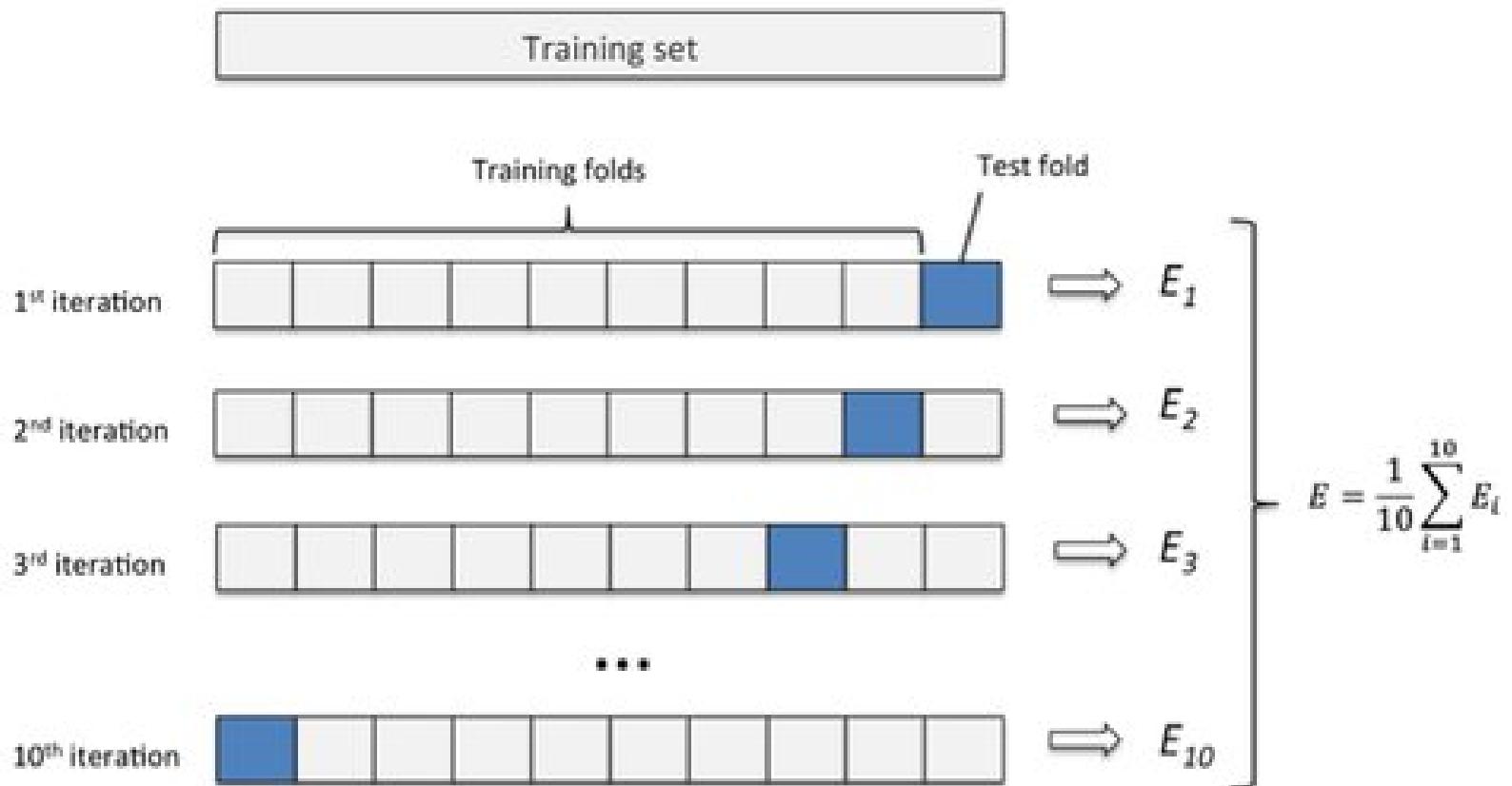
# Resampling procedures

- Cross-validation (CV) and bootstrapping are the two most popular classes of resampling methods in model building
- In model building, resampling is often used to
- assess the stability of a model
- infer on model parameters (tests, confidence intervals, point estimates, bias)
- measure prediction error

# Cross-validation algorithms

- Cross-validation schemes can be implemented in most statistical frameworks and for most estimation procedures
- The key principle of CV is to split the data into a training set and a validation set
- Estimators are built on the training set
- Validation set is used to validate/assess the performance of the estimators, e.g. estimating their prediction risk
- This training / validation splitting is repeated several times

# 10-fold cross validation



# Cross-validation algorithms

The most popular **cross-validation schemes** are,

- **Hold-out CV**, based on a single split of the data
- **$k$ -fold CV**: the data is split into  $k$  subsamples. Each subsample is successively removed for validation, the remaining data being used for training
- **Leave-one-out** (LOOCV) which corresponds to  **$n$ -fold CV**
- **Leave-  $q$ -out** every of the  $\binom{n}{q}$  possible subsets is removed for validation

# CV prediction error

- In  $k$ -fold CV the data is split into  $k$  subsamples (the  $k$  folds)
- Each subsample is successively removed for **validation**, the remaining data being used for **training**
- Thus, for each of the  $k$  folds  $f = 1, \dots, k$ ,  $\iota_f \subset \{1, \dots, n\}$  denotes the  $n_f = |\iota_f|$  observations in the  $f$ th fold and we calculate

$$\text{CV}_{(k)} = \frac{1}{k} \sum_{f=1}^k \text{MSE}_f = \frac{1}{k} \sum_{f=1}^k \frac{1}{n_f} \sum_{i \in \iota_f} [y_i - \hat{y}_{\alpha i}(-\iota_f)]^2$$

- Similarly, for estimating classification errors

# Choice of $k$ affects the quality of the CV error

- If  $k = 2$ : split-sample CV - CV error estimates **biased** upwards because only half the data used in the training samples
- If  $k = n$ : LOOCV - CV estimates have **high variance** because in the above equation a total of  $n$  positively correlated quantities are averaged;
- **Standard choices** are  $k = 10$  (default in `cv.glmnet` through option `nfold=10`) or  $k = 5$ : This balances bias and variance
- The  $k$  partly overlapping training sets can be used to learn more about the stability of selected models

# Case study: selecting $\lambda$ for Lasso

# How to best choose $\lambda$ ?

- Many options!
- Use information criteria along the lasso, ridge, or elastic-net path
- Use cross-validation
- as an alternative to `lambda.min` often `lambda.1se` does give better performance in terms of prediction
- `lambda.1se` is the largest  $\lambda$  at which the MSE is within one standard error of the minimal MSE (at `lambda.min`)

# Diabetes example

```
data("diabetes", package = "lars")
library("glmnet")
x = diabetes$x2 # includes squares and interactions
y = diabetes$y
lasso.fit = glmnet(x, y)
length(lasso.fit$lambda)
```

```
## [1] 100
```

```
lasso.predict = predict(lasso.fit, newx = x)
dim(lasso.predict)
```

```
## [1] 442 100
```

# Diabetes example: AIC to select $\lambda$

```
lasso.aic = rep(0, 100)
sigma.hat = summary(lm(y ~ x))$s
for (m in 1:100) {
  yhat = lasso.predict[, m]
  k = lasso.fit$df[m]
  lasso.aic[m] = sum((y - yhat)^2)/(sigma.hat^2) + 2*k
}
which.min(lasso.aic)
```

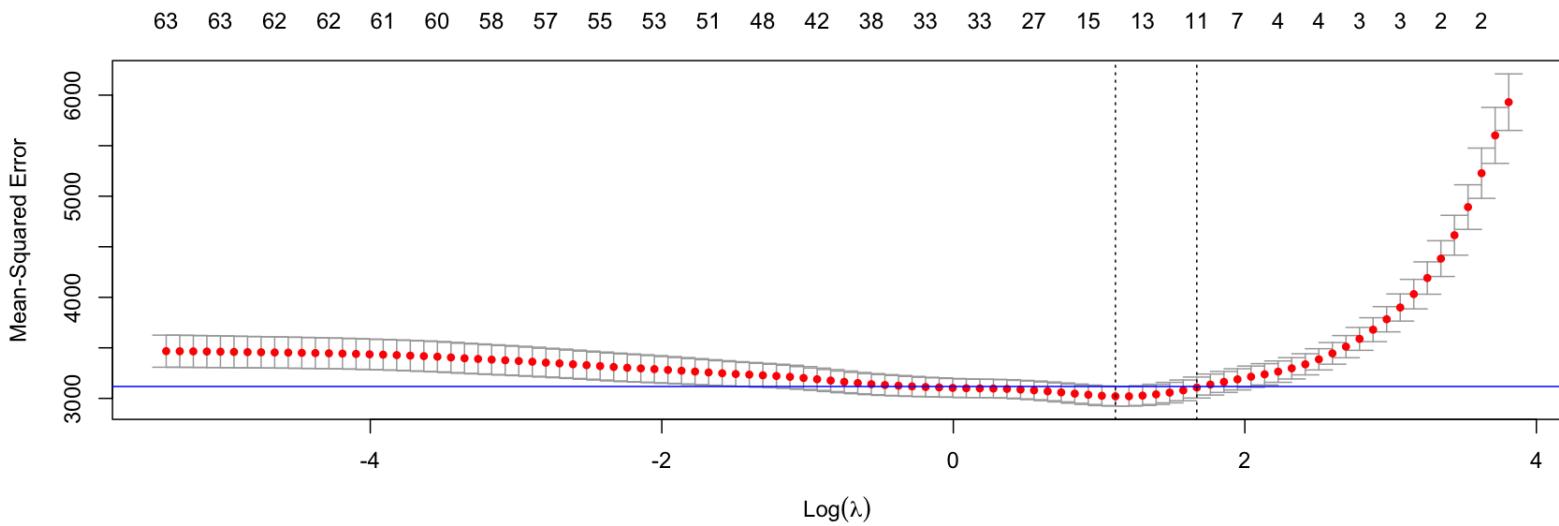
```
## [1] 32
```

```
lasso.fit$lambda[32]
```

```
## [1] 2.524812
```

# Diabetes example: CV to select $\lambda$

```
set.seed(2020)
lasso.cv = cv.glmnet(x, y)
plot(lasso.cv)
abline(h = min(lasso.cv$cvm) + lasso.cv$cvsd[which.min(lasso.cv$cvm)],
       col = "blue")
```



```
lasso.cv$lambda.1se
```

# Diabetes example:

Predictors selected by **AIC**:

```
aic_betas = glmnet(x, y, lambda = lasso.fit$lambda[32])$beta  
as.matrix(aic_betas)[as.matrix(aic_betas)>0, ]  
  
##          bmi         map        ltg        glu      age^2      bmi^2  
## 500.775931 262.976223 469.918325 25.416346 17.558198 42.899478  
##      glu^2    age:sex    age:map    age:ltg    age:glu    sex:map  
## 76.518096 115.463280 30.647942 12.662471  9.365541  7.745166  
##      bmi:map  
## 90.571069
```

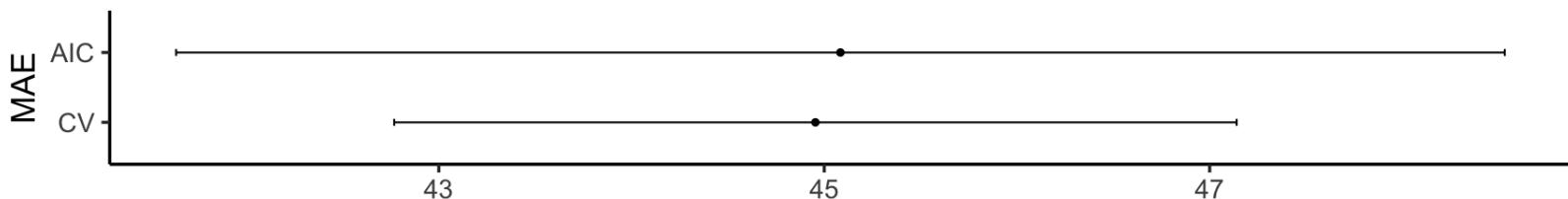
Predictors selected by **CV**:

```
cv_betas = glmnet(x, y, lambda = lasso.cv$lambda.1se)$beta  
as.matrix(cv_betas)[as.matrix(cv_betas)>0, ]  
  
##          bmi         map        ltg        bmi^2      glu^2    age:sex  
## 502.887614 205.892655 453.531965 17.266008 39.776736 60.581213  
##      age:map    age:glu    bmi:map  
## 14.945884   7.108415  59.023875
```

# Diabetes example

We can choose between the two competing models using **cross validation**.

```
library(caret)
control <- trainControl(method = "cv", number = 10)
dat <- cbind(y, x)
aic_cv <- train(y ~ bmi + map + ltg + glu + `age^2` + `bmi^2` +
                 `glu^2` + `age:sex` + `age:map` + `age:ltg` +
                 `age:glu` + `sex:map` + `bmi:map`,
                 method = "lm", data = dat, trControl = control)
cv_cv <- train(y ~ bmi + map + ltg + `bmi^2` + `glu^2` +
                 `age:sex` + `age:map` + `age:glu` + `bmi:map`,
                 method = "lm", data = dat, trControl = control)
results <- resamples(list(AIC = aic_cv, CV = cv_cv))
ggplot(results) + theme_classic(base_size = 18) +
  labs(x = "MAE")
```



## B2. The mplot package

# Installing the mplot package

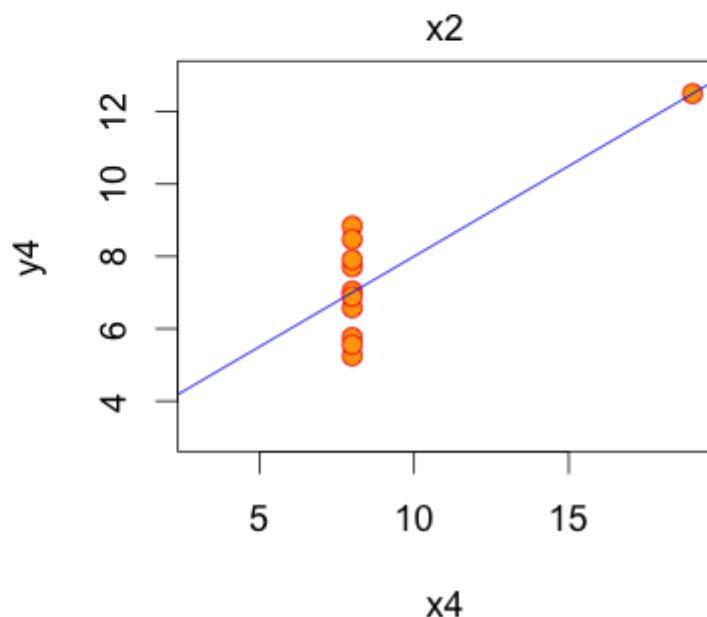
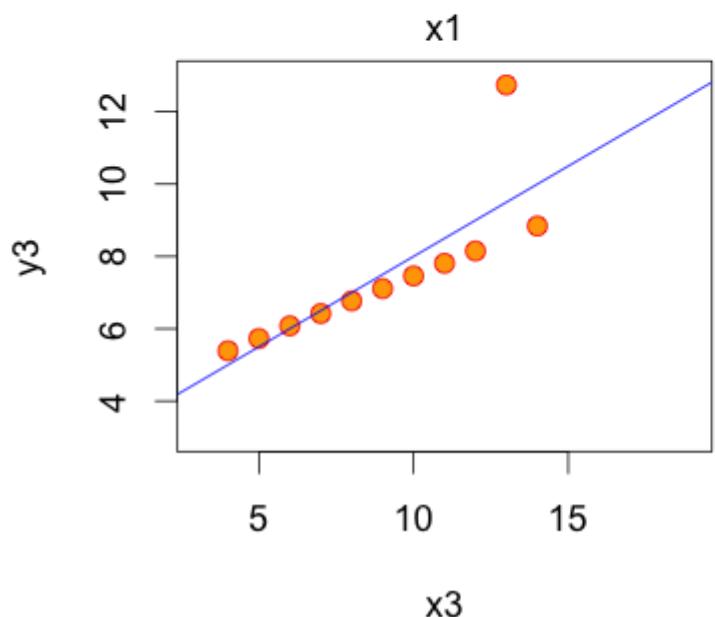
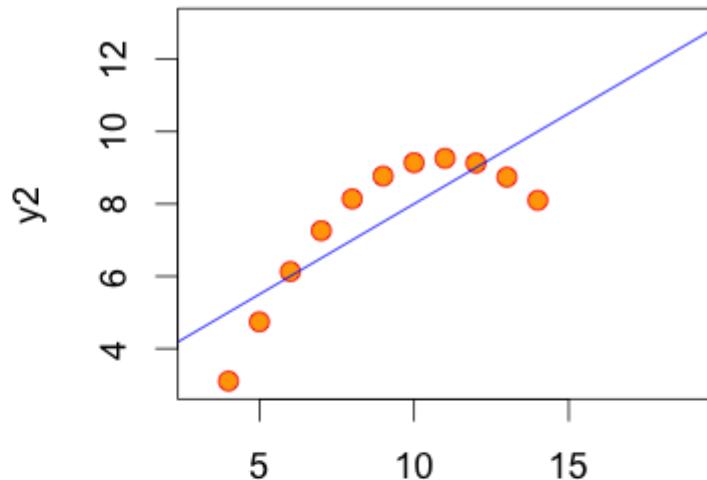
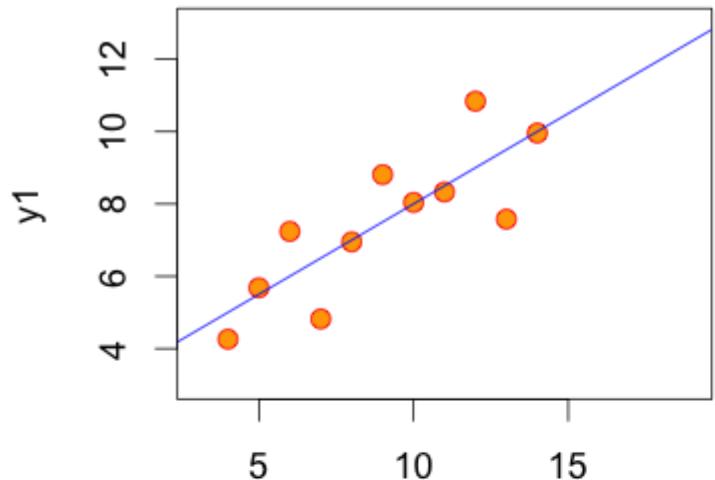
```
# install.packages("devtools")
# devtools::install_github("garthtarr/mplot")
install.packages("mplot")
library(mplot)
```

Website: <http://garthtarr.github.io/mplot>

## Main functions

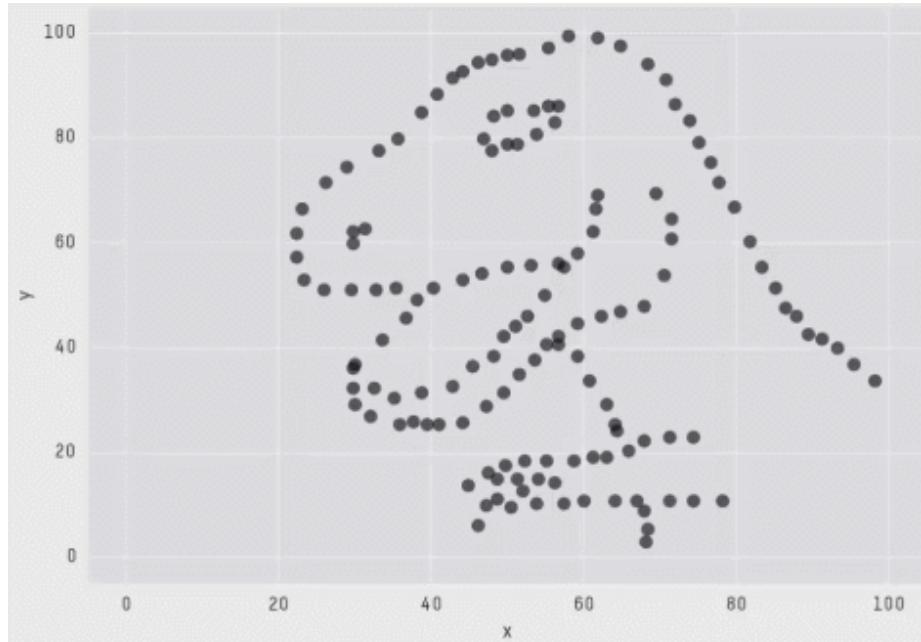
- `af()` for the adaptive fence for `lm()` and `glm()`
- `vis()` for variable importance plots and model stability plots for `lm()` and `glm()`
- `bglmnet()` for bootstrapping `glmnet()` and variable importance and model stability plots
- `mplot()` for an interactive shiny interface

# Motivation – Recall Anscombe's quartet



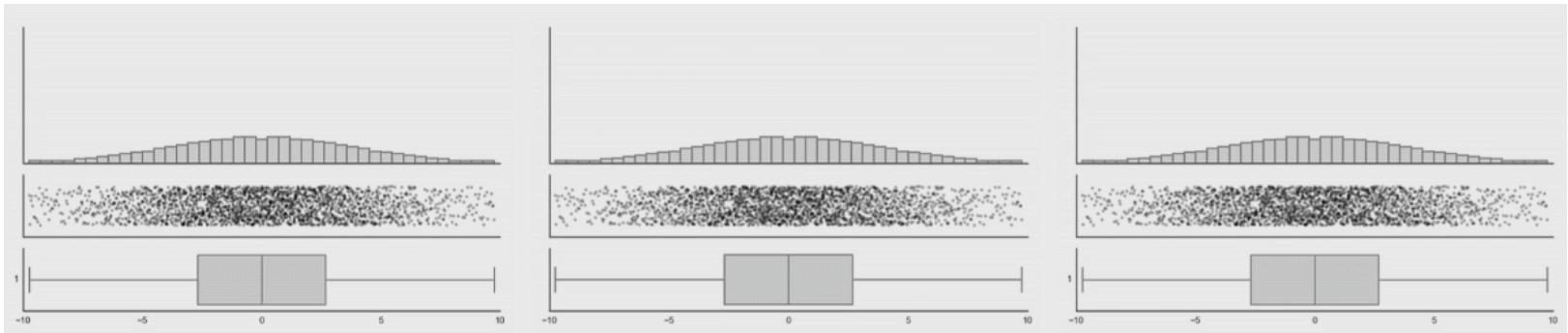
# Datasaurus

```
library(datasauRus)
```

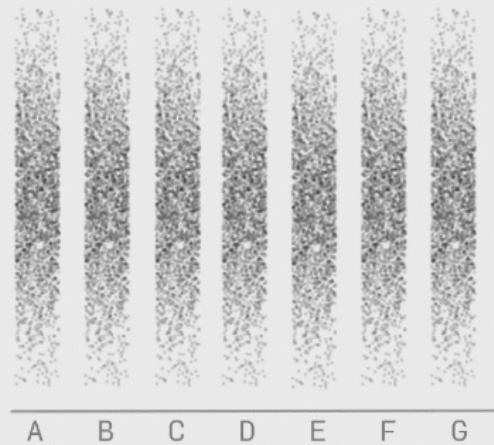


X Mean: 54.2659224  
Y Mean: 47.8313999  
X SD : 16.7649829  
Y SD : 26.9342120  
Corr. : -0.0642526

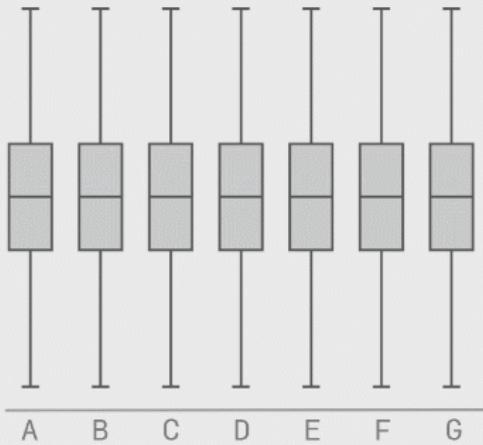
# See also



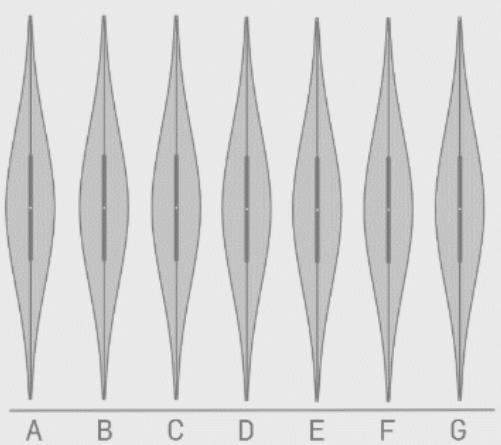
**Raw Data**



**Box-plot of the Data**



**Violin-plot of the Data**

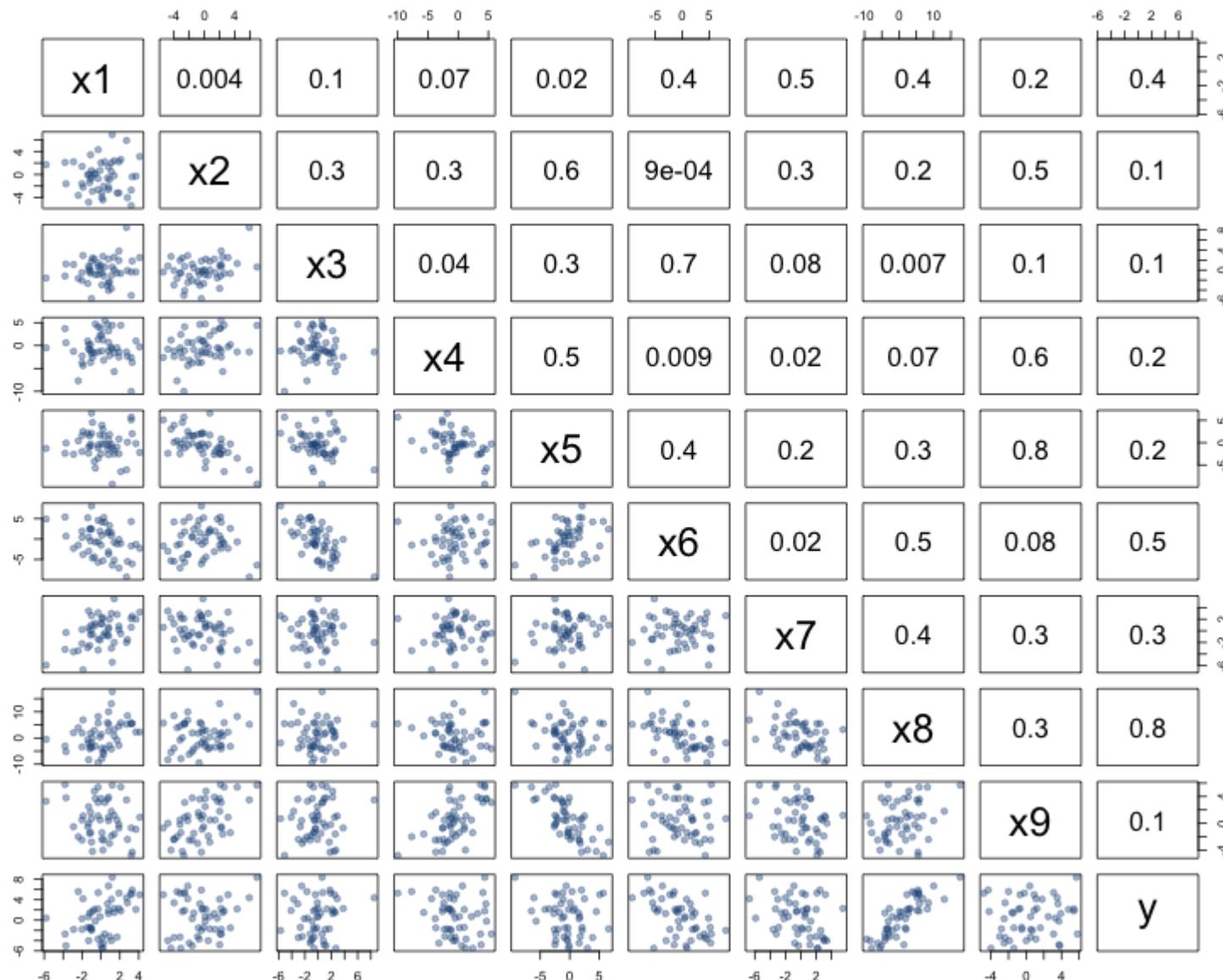


# Graphical tools for model selection

- Underused!
- Fence and its variants (series of papers by Jiang et al, starting with Jiang, Rao, Gu, and Nguyen (2008))
- Stability selection Meinshausen and Bühlmann (2010)
- Model selection curves Müller and Welsh (2010)
- Variable inclusion plots Müller and Welsh (2010) and Murray, Heritier, and Müller (2013)
- Variable selection with `mplot` (this lecture)

# Artificial example

# Artificial example: Correlations



# Artificial example: Full model

```
library(mplot)
lm.art = lm(y ~ ., data = artificial)
summary(lm.art)

##
## Call:
## lm(formula = y ~ ., data = artificial)
##
## Residuals:
##     Min      1Q  Median      3Q     Max 
## -4.7254 -1.2126  0.2456  1.3446  3.4808 
##
## Coefficients:
##             Estimate Std. Error t value Pr(>|t|)    
## (Intercept) -0.1002    0.3268  -0.307   0.761    
## x1           0.6403    0.6924   0.925   0.361    
## x2           0.2574    0.6176   0.417   0.679    
## x3          -0.5105    1.2391  -0.412   0.683    
## x4          -0.2978    0.2515  -1.184   0.243    
## x5           0.3551    0.5982   0.594   0.556    
## x6          -0.5428    0.9612  -0.565   0.575    
## x7          -0.4280    0.6308  -0.678   0.501    
## x8           0.1503    0.6175   0.243   0.809
```

# Artificial example: Final model

```
step(lm.art)

## Start: AIC=79.3
## y ~ x1 + x2 + x3 + x4 + x5 + x6 + x7 + x8 + x9
##
##          Df  Sum of Sq    RSS    AIC
## - x8      1    0.2423 163.94 77.374
## - x3      1    0.6946 164.39 77.512
## - x2      1    0.7107 164.41 77.517
## - x6      1    1.3051 165.00 77.698
## - x5      1    1.4425 165.14 77.739
## - x9      1    1.6065 165.31 77.789
## - x7      1    1.8835 165.58 77.873
## - x1      1    3.4999 167.20 78.358
## - x4      1    5.7367 169.44 79.023
## <none>           163.70 79.301
##
## Step: AIC=77.37
## y ~ x1 + x2 + x3 + x4 + x5 + x6 + x7 + x9
##
##          Df  Sum of Sq    RSS    AIC
## <none>           163.94 77.374
## - x2      1    20.259 184.20 81.227
```

# Artificial example: Final model

```
lm.fin = lm(y ~ . - x8, data = artificialeg)
summary(lm.fin)
```

```
## 
## Call:
## lm(formula = y ~ . - x8, data = artificialeg)
## 
## Residuals:
##      Min       1Q   Median       3Q      Max 
## -4.6881 -1.2352  0.2787  1.4102  3.5420 
## 
## Coefficients:
##             Estimate Std. Error t value Pr(>|t|)    
## (Intercept) -0.1143    0.3179  -0.360  0.720952    
## x1           0.8019    0.1941   4.132  0.000173 ***  
## x2           0.4011    0.1778   2.256  0.029438 *    
## x3          -0.8083    0.1914  -4.223  0.000131 ***  
## x4          -0.3514    0.1196  -2.938  0.005408 **   
## x5           0.4927    0.1933   2.548  0.014667 *    
## x6          -0.7738    0.1490  -5.194   6e-06 ***  
## x7          -0.5772    0.1465  -3.941  0.000309 ***  
## x9           0.5478    0.1890   2.899  0.005987 **  
## ---
```

# Artificial example: Data generating model

The true data generating model is:  $y = 0.6 x_8 + \varepsilon$ .

```
art.true = lm(y ~ x8, data = artificialeg)
summary(art.true)
```

```
##
## Call:
## lm(formula = y ~ x8, data = artificialeg)
##
## Residuals:
##     Min      1Q  Median      3Q     Max 
## -5.6013 -1.2985  0.1303  1.4605  3.8056 
##
## Coefficients:
##             Estimate Std. Error t value Pr(>|t|)    
## (Intercept) 0.03235   0.29458   0.11    0.913    
## x8          0.54928   0.05264  10.43 6.18e-14 ***  
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Residual standard error: 2.046 on 48 degrees of freedom
## Multiple R-squared:  0.694,    Adjusted R-squared:  0.6877 
## F-statistic: 108.9 on 1 and 48 DF,  p-value: 6.18e-14
```

# Information criteria can be blind

Use resampling and graphs to **see** more!

# Bootstrapping regression models

The **bootstrap** is a resampling strategy to generate  $B$  training sets

## Popular implementations

- **Paired bootstrap**: samples (with replacement) rows of  $[\mathbf{y}, \mathbf{X}]$
- **Weighted bootstrap** (used in `vis()` and `bglmnet()`): mimics the paired bootstrap by keeping  $[\mathbf{y}, \mathbf{X}]$  as is but weighs each observation (row) through using a weights vector, where the weights are sampled from a distribution with expectation 1 (e.g. exponential).
- **Parametric bootstrap** (used in `af()`): estimate the (parametric) distribution of  $\mathbf{y}$  (or  $\boldsymbol{\varepsilon}$ ) and draw new observations from it
- **Residual bootstrap**: baseline model + bootstrapped error

# Stability

- In practice it is important that selected models are stable

**Key idea:** small changes should have small effects

- Example: Information criterion is *unstable* when  $\hat{\alpha}(\lambda)$  is selected with dimension  $p_{\hat{\alpha}(\lambda)}$  but, for some small  $\delta > 0$ ,  $\hat{\alpha}(\lambda + \delta)$  selected with  $p_{\hat{\alpha}(\lambda+\delta)} < p_{\hat{\alpha}(\lambda)}$
- Investigate model selection criterion in a neighborhood of  $\lambda$

# Obtaining stability information

**Use resampling:** repeatedly calculate how well (the best) model(s) perform given  $\lambda$ , summarise performance with statistics and graphs

We will do the following:

- Investigate model detectability as a function of  $\lambda$
- Estimate probability that model  $\alpha \in \mathcal{A}$  is optimal at  $\lambda$
- Estimate best model(s) within each size/dimension
- Show model selection curves also for models that are never optimal

# Variable inclusion plots

# Variable inclusion plots

**Aim:** To visualise inclusion probabilities as a function of the penalty multiplier  $\lambda \in [0, 2 \log(n)]$

## Procedure

1. Calculate (weighted) **bootstrap samples**  $b = 1, \dots, B$ .
2. For each **bootstrap sample**, at each  $\lambda$  value, find  $\hat{\alpha}_{\lambda}^{(b)} \in \mathcal{A}$  as the model with smallest  $\text{GIC}(\alpha; \lambda) = -2 \times \text{LogLik}(\alpha) + \lambda p_{\alpha}$ .
3. The **inclusion probability** for variable  $x_j$  is estimated as  
$$\frac{1}{B} \sum_{b=1}^B \mathbf{1}\{j \in \hat{\alpha}_{\lambda}^{(b)}\}.$$

The **inclusion probability** for variable  $x_j$  is the proportion of times variable  $x_j$  is in the best model

- Müller and Welsh (2010) for linear regression models
- Murray, Heritier, and Müller (2013) for generalised linear models

# Artificial example: VIP

```
library(mplot)
vis.art = vis(lm.art)
plot(vis.art, which = "vip", interactive = TRUE)
```

# Model stability plots

# Artificial example: Loss against size

```
plot(vis.art, which="lvk", interactive = TRUE)
```

# Artificial example: Loss against size

```
plot(vis.art, which = "lvk", highlight = "x6", interactive = TRUE)
```

# Model stability plots

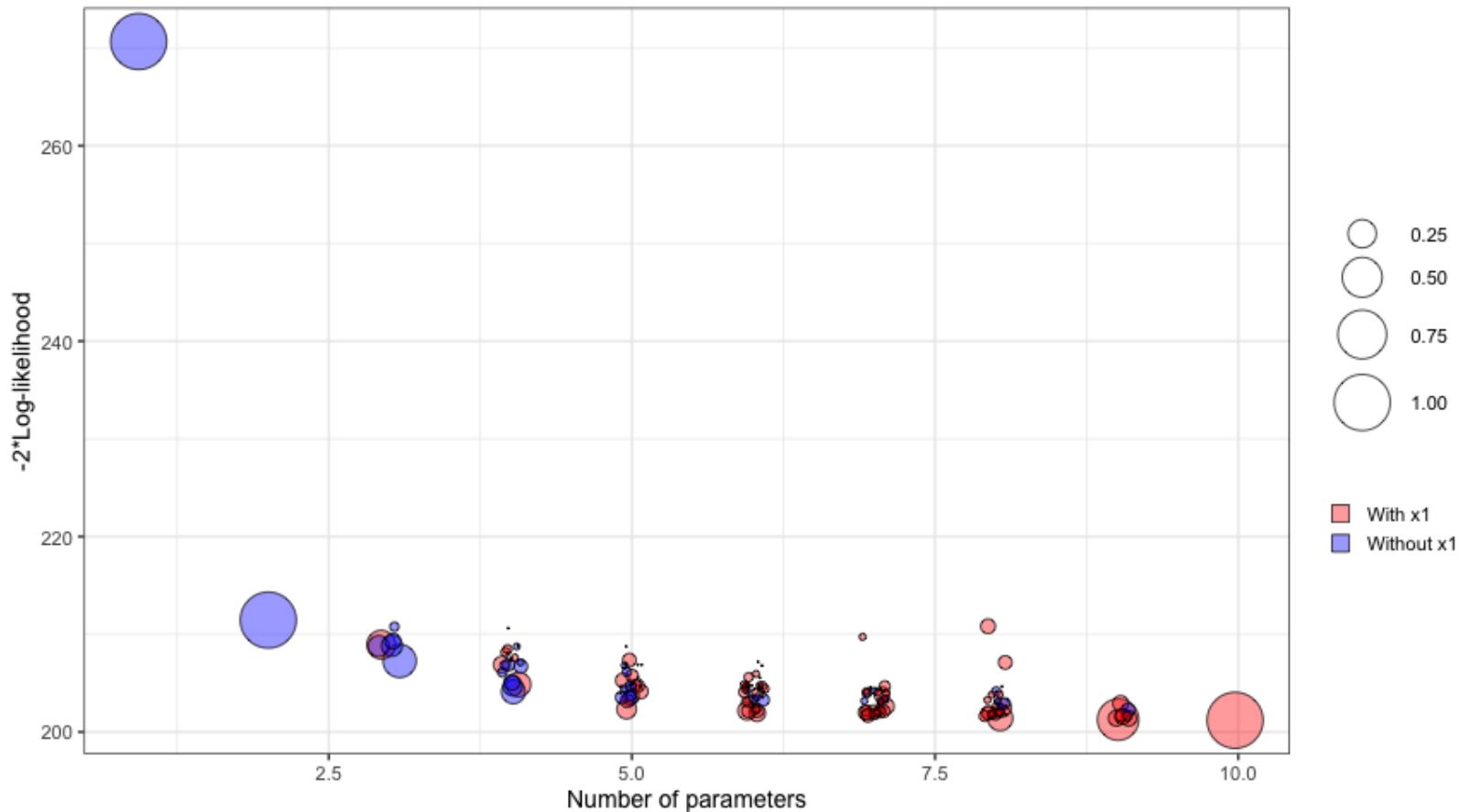
**Aim:** To add value to the loss against size plots by choosing a symbol size proportional to a measure of stability

## Procedure

1. Calculate (weighted) bootstrap samples  $b = 1, \dots, B$
2. For each bootstrap sample, identify the *best* model at each dimension
3. Add this information to the loss against size plot using model identifiers that are proportional to the frequency with which a model was identified as being *best* at each model size

# Artificial example: Model stability plot

```
plot(vis.art, which = "boot", interactive = FALSE)
```



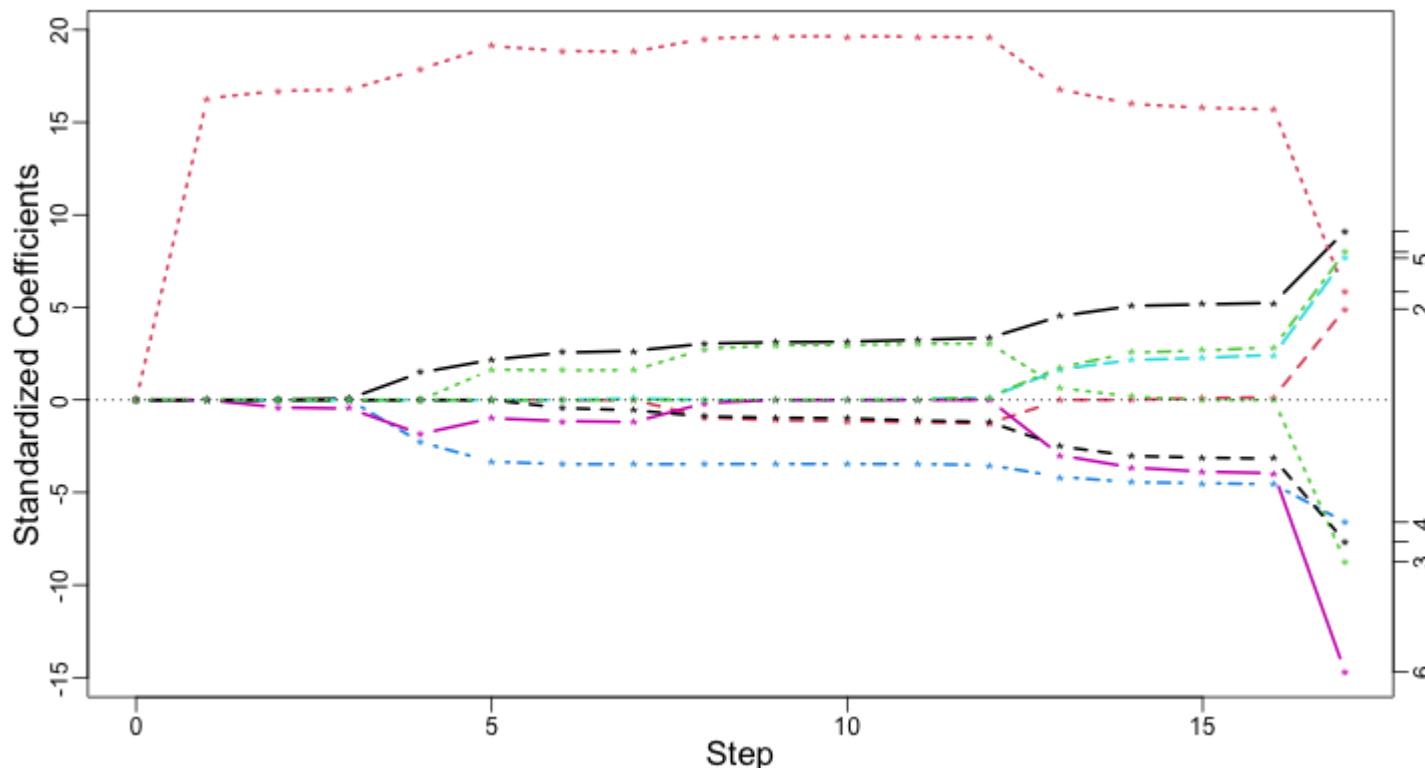
# Artificial example: Model stability plot

```
plot(vis.art, which = "boot", interactive = TRUE)
```

# Bootstrapping the lasso

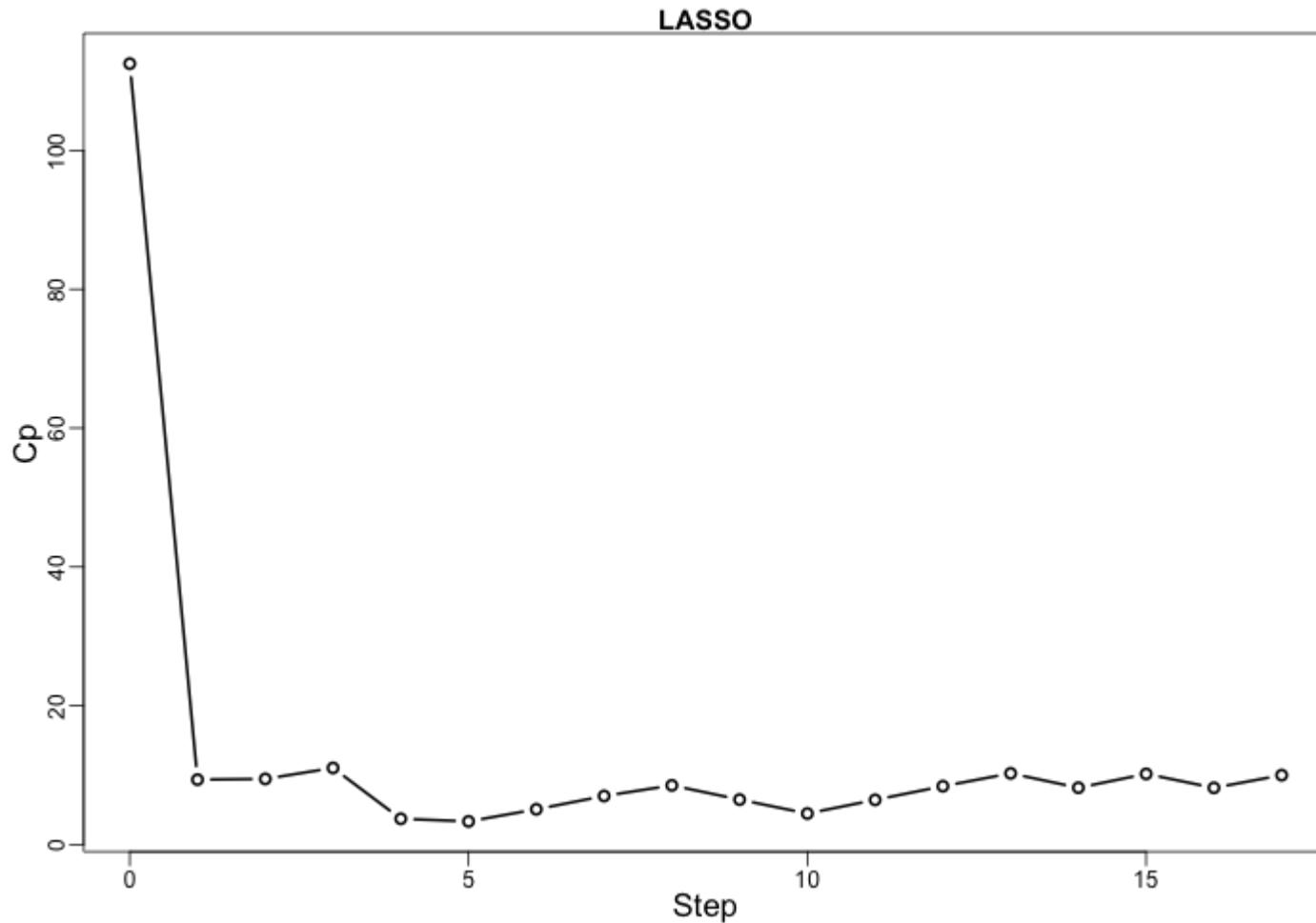
# Artificial example: Lasso

```
library(lars)
x = as.matrix(subset(artificialeg, select = -y))
y = as.matrix(subset(artificialeg, select = y))
art.lars=lars(x, y)
plot(art.lars, xvar = "step", breaks = FALSE, lwd = 2, cex.lab = 1.4)
```

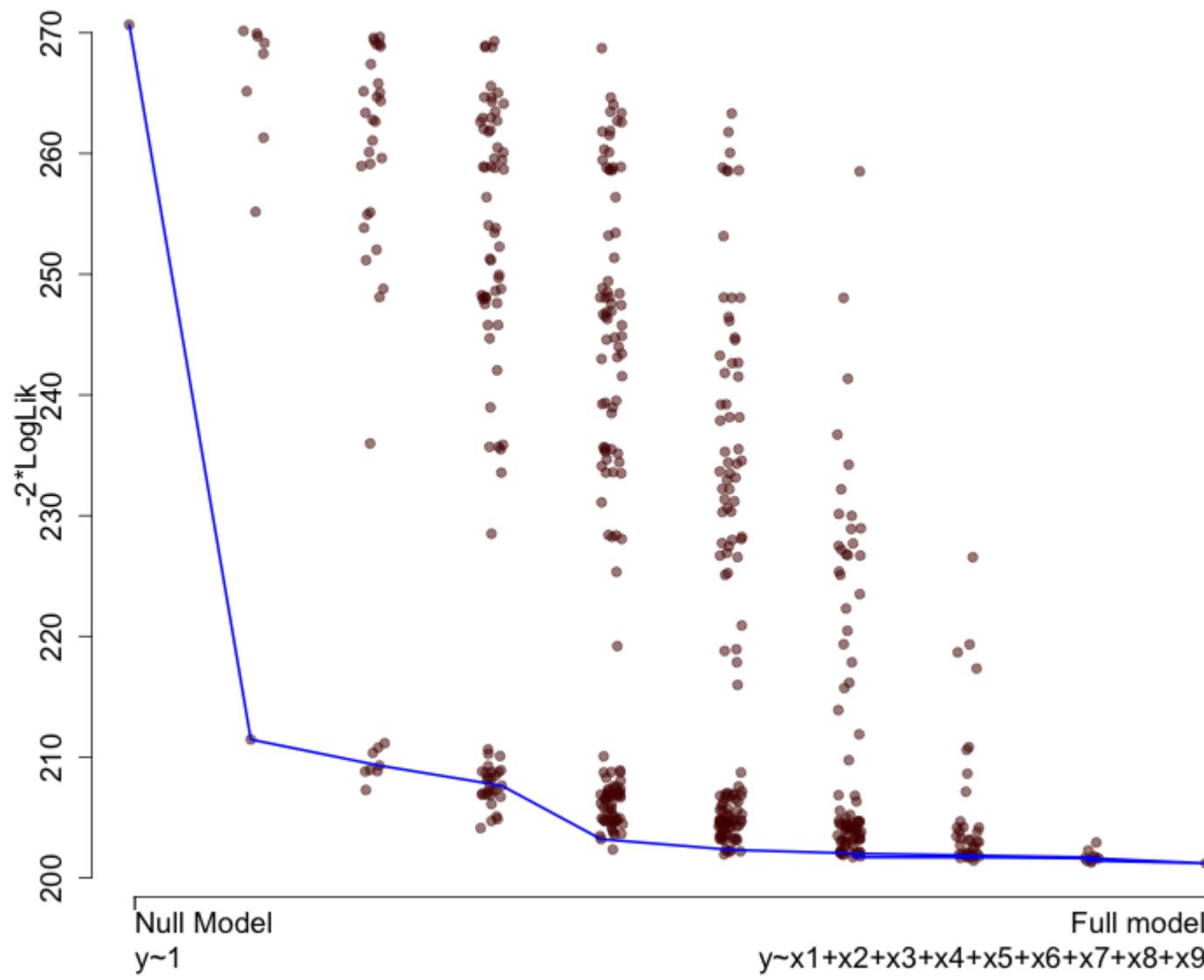


# Artificial example: Lasso

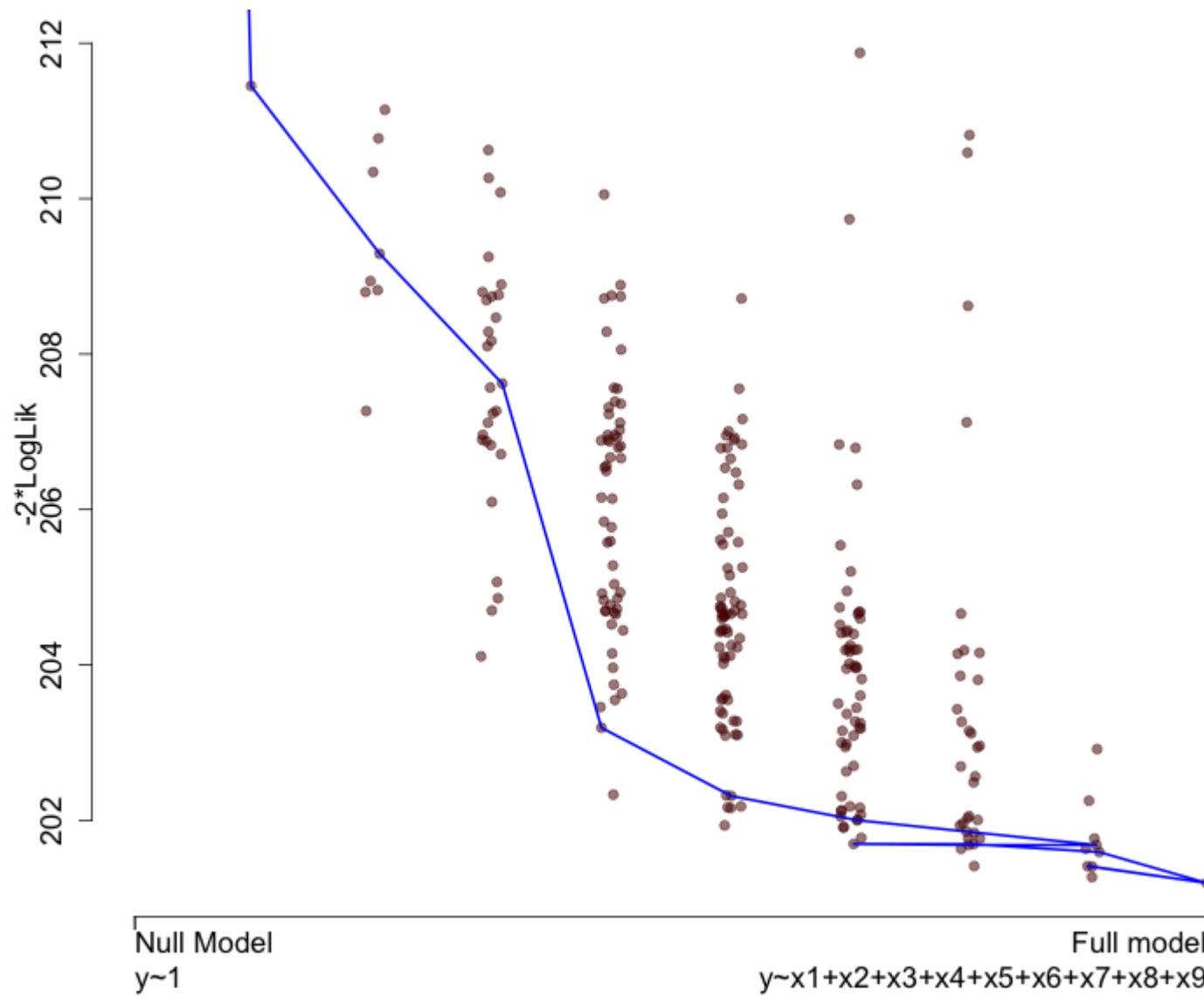
```
plot(art.lars, xvar = "step", plottype = "Cp", lwd = 2, cex.lab = 1.4)
```



# Artificial example: Lasso



# Artificial example: Lasso (zoomed)



# Bootstrapping the lasso

```
bgn.art = bgglmnet(lm.art, lambda = seq(0.05, 2.75, 0.1))
plot(bgn.art, which = "boot", interactive = TRUE)
```

# Bootstrapping the lasso

```
bgn.art = bglmnet(lm.art)
plot(bgn.art, which = "vip", interactive = TRUE)
```

## B3. Subtractive stability measures

$$\hat{Q}(M) = s - \sum_{j \in M} s_j$$

# Stability measures

- Use **resampling**, say  $B$  resamples (100, 1000 or more)
- Consider **baseline models** (e.g. all models on a solution path), say  $K$  such models
- Learn from an array of  $K \times p \times B$  regression coefficients
- Calculate for each feature  $j = 1, \dots, p$
- Inclusion frequency
- Exclusion frequency
- **Other variable importance measure**

# What is a subtractive measure?

Ingenious idea by Jiang, Nguyen and Rao (2011, Statistics and Its Interface)

A **subtractive measure**  $\hat{Q}$  satisfies

$$\hat{Q}(M_K) = s - \sum_{j \in M_K} s_j$$

with  $s_j \geq 0, k = 1, \dots, K$ .

Think "**lack-of-fit**" measure (like a log-likelihood) with "**benefits**" (something additional to a log-likelihood).

# Toy example

- Let the full model be  $\{1, 2, 3, 4, 5, 6, 7, 8, 9, 10\}$ , i.e.  $p = 10$
- Let's assume that  $s_1 = |-0.05|$ ,  $s_2 = |-0.19|$ , with remaining  $s_j$ 's being 0.34, 0.21, 0.30, 0.19, 0.02, 0.09, **0.42** and 0.03
- Note: all  $s_j$  values are non-negative
- Let  $s = \sum s_j = 1.84$
- That model minimizing  $\hat{Q}$  over all models with one variable is  $\{9\}$
- The best model with two variables is  $\{3, 9\}$
- $\hat{Q} \geq 0$  by construction, with equality for the full model

# What are the benefits?

Magic in  $s_j$  and constructing  $\hat{Q}$

- Fast access to identify best model of size  $d$
- Remains fast when bootstrapping

Full model fit sufficient to get well performing  $s_j$ 's

- $s_j = |\hat{\beta}_j|$  (**robust, penalised**)
- $s_j = |t_j|$  (standardised coefficients)
- **Robust quasi-deviance statistics** (differences to full/large baseline model)

# Coming soon to the mplot package

- Resample  $\{y, \mathbf{X}\}$ , e.g. through weighted bootstrap
- Use OLS (LIMMA), Ridge, Lasso, Elastic-net, etc
- Observe  $s_j^{(b)}$ ,  $b = 1, \dots, B$ ,  $j = 1, \dots, p$
- Find the empirical probability that  $s_j > s_k$
- Derive new indices
- Mean bootstrapped rank of  $s_j^{(b)}$
- Standard deviation of  $\text{rank}(s_j^{(b)})$
- Devise new visualisations
- Preprints: Smith, Günnewig, and Müller (2020) and Smith, Müller, and Günnewig (2020)

# A function to bootstrap the full model

```
sstab = function(mf, B = 100) {  
  full_coeff = coefficients(mf)  
  kf = length(full_coeff)  
  coef.res = matrix(ncol = kf, nrow = B)  
  colnames(coef.res) = names(full_coeff)  
  formula = stats::formula(mf)  
  data = model.frame(mf)  
  for(i in 1:B){  
    wts = stats::rexp(n = length(resid(mf)), rate = 1)  
    data$wts = wts  
    mod = stats::lm(formula = formula,  
                    data = data,  
                    weights = wts)  
    coef.res[i, ] = coefficients(mod)  
  }  
  return(coef.res)  
}
```

# Diabetes example

```
data("diabetes", package = "lars")
x = diabetes$x
y = diabetes$y
df = data.frame(scale(cbind(y, x)))
lm1 = lm(y ~ ., data = df)
sj = sstab(lm1)
round(head(sj[, -1], n = 10), 2)
```

```
##          age    sex   bmi   map     tc    ldl    hdl    tch    ltg    glu
## [1,] -0.05 -0.19  0.34  0.21 -0.30  0.19 -0.02  0.09  0.42  0.03
## [2,]  0.02 -0.16  0.29  0.23 -0.21 -0.04  0.08  0.29  0.29  0.02
## [3,]  0.02 -0.04  0.26  0.17 -0.37  0.20  0.05  0.20  0.43  0.06
## [4,]  0.00 -0.15  0.32  0.26 -0.39  0.11  0.04  0.16  0.40  0.01
## [5,] -0.04 -0.17  0.34  0.21 -0.56  0.30  0.10  0.15  0.50  0.06
## [6,] -0.05 -0.11  0.29  0.18 -0.64  0.47  0.11  0.06  0.50  0.08
## [7,]  0.01 -0.17  0.28  0.23 -0.31  0.03 -0.05  0.14  0.41  0.08
## [8,]  0.00 -0.12  0.37  0.15 -0.24  0.06 -0.09  0.06  0.34  0.07
## [9,]  0.00 -0.04  0.33  0.14 -0.40  0.20  0.11  0.08  0.50  0.08
## [10,] 0.01 -0.17  0.34  0.24 -0.76  0.48  0.17  0.15  0.56 -0.01
```

# Diabetes example

```
sj = abs(sj[, -1])
sj_ranks = apply(sj, 1, rank)
head(t(sj_ranks), n = 10)
```

```
##      age sex bmi map tc ldl hdl tch ltg glu
## [1,]   3   6   9   7   8   5   1   4   10  2
## [2,]   2   5   9   7   6   3   4   8   10  1
## [3,]   1   2   8   5   9   7   3   6   10  4
## [4,]   1   5   8   7   9   4   3   6   10  2
## [5,]   1   5   8   6  10   7   3   4   9   2
## [6,]   1   5   7   6  10   8   4   2   9   3
## [7,]   1   6   8   7   9   2   3   5   10  4
## [8,]   1   6  10   7   8   2   5   3   9   4
## [9,]   1   2   8   6   9   7   5   3   10  4
## [10,]  1   4   7   6  10   8   5   3   9   2
```

# Diabetes example

```
sj_rank_mean = sort(apply(sj_ranks, 1, mean), decreasing = TRUE)
sj_rank_mean
```

```
##   ltg    tc    bmi   map   ldl   sex   tch   hdl   glu   age
## 9.33 8.81 8.06 6.21 6.21 4.67 4.33 3.40 2.38 1.60
```

```
# tc
table(sj_ranks[5, ])/100
```

```
##
##      1      3      4      5      6      7      8      9      10
## 0.02 0.01 0.02 0.02 0.02 0.02 0.02 0.18 0.22 0.49
```

```
# ltg
table(sj_ranks[9, ])/100
```

```
##
##      6      8      9     10
## 0.01 0.03 0.57 0.39
```

# References

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```
devtools::session_info(include_base = FALSE)
```

## - Session info	## httr	1.4.1	2019-08-05 [1] CRAN (R 4.0.0)
## setting value	## ipred	0.9-9	2019-04-28 [1] CRAN (R 4.0.0)
## version R version 4.0.0 (2020-04-24)	## iterators	1.0.12	2019-07-26 [1] CRAN (R 4.0.0)
## os macOS Catalina 10.15.5	## jsonlite	1.7.0	2020-06-25 [1] CRAN (R 4.0.0)
## system x86_64, darwin17.0	## knitr	1.29	2019-09-15 [1] CRAN (R 4.0.0)
## ui X11	## labeling	0.3	2020-06-23 [1] CRAN (R 4.0.0)
## language (EN)	## lars	* 1.2	2014-08-23 [1] CRAN (R 4.0.0)
## collate en_AU.UTF-8	## later	1.1.0.1	2013-04-24 [1] CRAN (R 4.0.0)
## ctype en_AU.UTF-8	## lattice	* 0.20-41	2020-06-05 [1] CRAN (R 4.0.0)
## tz Australia/Sydney	## lava	1.6.7	2020-04-02 [1] CRAN (R 4.0.0)
## date 2020-07-16	## leaps	3.1	2020-03-05 [1] CRAN (R 4.0.0)
##	## lifecycle	0.2.0	2020-01-16 [1] CRAN (R 4.0.0)
## - Packages	## lubridate	1.7.9	2020-03-06 [1] CRAN (R 4.0.0)
## package * version date lib source	## magrittr	1.5	2014-11-22 [1] CRAN (R 4.0.0)
## assertthat 0.2.1 2019-03-21 [1] CRAN (R 4.0.0)	## MASS	7.3-51.6	2020-04-26 [1] CRAN (R 4.0.0)
## backports 1.1.8 2020-06-17 [1] CRAN (R 4.0.0)	## Matrix	* 1.2-18	2019-11-27 [1] CRAN (R 4.0.0)
## bibtex 0.4.2.2 2020-01-02 [1] CRAN (R 4.0.0)	## memoise	1.1.0	2017-04-21 [1] CRAN (R 4.0.0)
## callr 3.4.3 2020-03-28 [1] CRAN (R 4.0.0)	## mime	0.9	2020-02-04 [1] CRAN (R 4.0.0)
## caret * 6.0-86 2020-03-20 [1] CRAN (R 4.0.0)	## ModelMetrics	1.2.2.2	2020-03-17 [1] CRAN (R 4.0.0)
## class 7.3-17 2020-04-26 [1] CRAN (R 4.0.0)	## mplot	* 1.0.4	2020-02-15 [1] CRAN (R 4.0.0)
## cli 2.0.2 2020-02-28 [1] CRAN (R 4.0.0)	## munsell	0.5.0	2018-06-12 [1] CRAN (R 4.0.0)
## codetools 0.2-16 2018-12-24 [1] CRAN (R 4.0.0)	## nlme	3.1-148	2020-05-24 [1] CRAN (R 4.0.0)
## colorspace 1.4-1 2019-03-18 [1] CRAN (R 4.0.0)	## nnet	7.3-14	2020-04-26 [1] CRAN (R 4.0.0)
## crayon 1.3.4 2017-09-16 [1] CRAN (R 4.0.0)	## pillar	1.4.4	2020-05-05 [1] CRAN (R 4.0.0)
## data.table 1.12.8 2019-12-09 [1] CRAN (R 4.0.0)	## pkgbuild	1.0.8	2020-05-07 [1] CRAN (R 4.0.0)
## datasauRus * 0.1.4 2018-09-20 [1] CRAN (R 4.0.0)	## pkgconfig	2.0.3	2019-09-22 [1] CRAN (R 4.0.0)
## desc 1.2.0 2018-05-01 [1] CRAN (R 4.0.0)	## pkgload	1.1.0	2020-05-29 [1] CRAN (R 4.0.0)
## devtools 2.3.0 2020-04-10 [1] CRAN (R 4.0.0)	## plyr	1.8.6	2020-03-03 [1] CRAN (R 4.0.0)
## digest 0.6.25 2020-02-23 [1] CRAN (R 4.0.0)	## prettyunits	1.1.1	2020-01-24 [1] CRAN (R 4.0.0)
## doParallel 1.0.15 2019-08-02 [1] CRAN (R 4.0.0)	## pROC	1.16.2	2020-03-19 [1] CRAN (R 4.0.0)
## doRNG 1.8.2 2020-01-27 [1] CRAN (R 4.0.0)	## processx	3.4.2	2020-02-09 [1] CRAN (R 4.0.0)
## dplyr 1.0.0 2020-05-29 [1] CRAN (R 4.0.0)	## prodlim	2019.11.13	2019-11-17 [1] CRAN (R 4.0.0)
## ellipsis 0.3.1 2020-05-15 [1] CRAN (R 4.0.0)	## promises	1.1.1	2020-06-09 [1] CRAN (R 4.0.0)
## evaluate 0.14 2019-05-28 [1] CRAN (R 4.0.0)	## ps	1.3.3	2020-05-08 [1] CRAN (R 4.0.0)
## fansi 0.4.1 2020-01-08 [1] CRAN (R 4.0.0)	## purrr	0.3.4	2020-04-17 [1] CRAN (R 4.0.0)
## farver 2.0.3 2020-01-16 [1] CRAN (R 4.0.0)	## R6	2.4.1	2019-11-12 [1] CRAN (R 4.0.0)
## fastmap 1.0.1 2019-10-08 [1] CRAN (R 4.0.0)	## Rcpp	1.0.4.6	2020-04-09 [1] CRAN (R 4.0.0)
## foreach 1.5.0 2020-03-30 [1] CRAN (R 4.0.0)	## recipes	0.1.13	2020-06-23 [1] CRAN (R 4.0.0)
## fs 1.4.1 2020-04-04 [1] CRAN (R 4.0.0)	## RefManageR	1.2.12	2019-04-03 [1] CRAN (R 4.0.0)
## generics 0.0.2 2018-11-29 [1] CRAN (R 4.0.0)	## remotes	2.1.1	2020-02-15 [1] CRAN (R 4.0.0)
## ggplot2 * 3.3.2 2020-06-19 [1] CRAN (R 4.0.0)	## reshape2	1.4.4	2020-04-09 [1] CRAN (R 4.0.0)
## glmnet * 4.0-2 2020-06-16 [1] CRAN (R 4.0.0)	## rlang	0.4.6	2020-05-02 [1] CRAN (R 4.0.0)
## glue 1.4.1 2020-05-13 [1] CRAN (R 4.0.0)	## rmarkdown	2.3	2020-06-18 [1] CRAN (R 4.0.0)
## googleVis 0.6.5 2020-06-08 [1] CRAN (R 4.0.0)	## rngtools	1.5	2020-01-23 [1] CRAN (R 4.0.0)
## gower 0.2.2 2020-06-23 [1] CRAN (R 4.0.0)	## rpart	4.1-15	2019-04-12 [1] CRAN (R 4.0.0)
## gtable 0.3.0 2019-03-25 [1] CRAN (R 4.0.0)	## rprojroot	1.3-2	2018-01-03 [1] CRAN (R 4.0.0)
## htmltools 0.5.0 2020-06-16 [1] CRAN (R 4.0.0)	## scales	1.1.1	2020-05-11 [1] CRAN (R 4.0.0)
## httpuv 1.5.4 2020-06-06 [1] CRAN (R 4.0.0)	## sessioninfo	1.1.1	2018-11-05 [1] CRAN (R 4.0.0)
	## shape	1.4.4	2018-02-07 [1] CRAN (R 4.0.0)
	## shiny	1.5.0	2020-06-23 [1] CRAN (R 4.0.0)
	## shinydashboard	0.7.1	2018-10-17 [1] CRAN (R 4.0.0)
	## stringi	1.4.6	2020-02-17 [1] CRAN (R 4.0.0)
	## stringr	1.4.0	2019-02-10 [1] CRAN (R 4.0.0)